THE DISTRIBUTION OF MOLYBERNUM
IN RED ALDER AND FIVE CTHER
NORTHWESTERN TREES

Bernard T. Bormann
The Evergreen State College
Olympia, Washington

ABSTRACT

Molybdenum, a necessary metallic coenzyme for nitrogen fixation, was measured in six northwest tree species. Concentrations in leaf-branch samples were lowest in Alnus rubra, a nitrogen fixing species, and highest in Pseudotsuga menziezii. The highest concentrations, measured in seeds of Alnus rubra, may be an adaptive mechanism allowing seedlings to rapidly begin nitrogen fixation. Alder frequently comes up on: cutover Douglas fir stands. The high concentration of Mo in Douglas fir litter may contribute to the success of these alder invasions.

INTRODUCTION

Of the twenty-four elements known to be essential to plant and microorganism growth, molybdenum is one of the eight so-called trace elements or micronutrients, and is required in very small amounts. Molybdenum nutrition of agricultural plants has justly received considerable attention, yet its utilization by forest ecosystems has redieved little study. This paper reports results of an initial study of the molybdenum distribution in various Facific northwest trees.

World-wide interest in rolybdenum began in 1942, when it was discovered that as little as 1/16 oz Mo per acre (4 g/ha) applied to unproductive clover pasture resulted in significant increases in clover production (anderson 1946). Subsequently molybdenum has been shown to be essential in nitrogen-fixing and other plants, and to be part of several enzymes including nitrate reductase and nitrogenase (Anderson 1956).

Perceived That a nitrogen deficiency.

The molybdenum content of most soils is low, averaging around 2 ppm, and is very uniform in its distribution (Rubins 1956). The availability of molybdenum for plant uptake is strongly influenced by soil pH. Below a pH of 5, molybdenum is almost completely bound to colloidal surfaces and becomes unavailable (Jones 1957).

The average molybdenum content of plant leaf tissue is

reported to be 0.10 ppm (Epstein 1965). It is known that some plants can highly concentrate specific elements, but no known accumulators of molybdenum have been found. Young and Guinn (1966), in one of the few intensive studies of micronutrient content of forest trees, found an average concentration of 3.5 ppm Mo in organs of seven tree species growing in Maine. Molybdenum occured in roots, bark, leaves and woody tissue in order of decreasing concentration.

To initiate a study of the role of molycdenum in the ecology of northwestern forests, two studies were carried out. In the first, concentrations of molybdenum were determined in six common northwest forest trees to look for differential distribution. In the second, various tissues of Alnus rubra were analyzed to determine if molybdenum is concentrated in specific organs.

Red alder (Alnus rubra Bongard) was chosen as the focal point of this study because of its importance in northwestern forest ecology and the possibility of its susceptability to molybdenum deficiency. It is a successful successional tree because of its ability to fix nitrogen, its very fast growth (as much as 5 ft./yr.) and its ability to tolerate and utilize high light intensities. It appears quickly on moist soils after conifers have been removed by fire or cutting. Aed alder has a large biomass of nodules and a high fixation efficiency of 814 ng N/g dry wt. nodules (Meyer 1966).

Zavitkovski and Newton (1968) reported a rate of over 300

**COMPART WITH STANDARD WITH STANDARD WITH STANDARD FOR Pigum satirum

and 258 mg N/g dry wt. nodules for Soja sp.

kg N fixed/ha yr in young dense stands. The average annual rate is probably 100-200 kg/ha.

METHODS

Samples were collected from a wooded area of The Evergreen State College campus, Clympia, washington. The soil in the area is a sandy loam with a pH range of 4-5.

Using special precautions, due to the nature of working with trace quantities, a forty year old red alder was felled and various organs sampled. Leaf-branch samples from alnus rubra, Tsuza heterophylla, Thuja plicata, Pseudotsuza menziezii, Taxus brevifolia and acer macrophyllum were also collected. Alder samples, except seeds, and all other samples, except Douglas fir, were collected and analyzed in May and June 1975. Local alder seed and Douglas fir samples were collected and analyzed in November 1975. An alder seed sample from Blue Lake, Humbolt Co., California was purchased and analyzed in June 1975.

In the lab these samples were partially dried and ground to a powder in a Wiley Mill and then fully dried overnight at 110°C, prior to being weighed for digestion.

The analytical technique utilized involves a perchloric acid digestion followed by a newly developed technique using graphite furnace atomic absorption and a standard addition method-ology to overcome interferences. A full description of this method can be obtained from the author.

Due to time required to perfect the technique and certain time limitations, only one analysis was conducted on each sample

except alder nodules. Some idea of the accuracy of the technique can be gained from two lines of evidence.

- 1.) Analysis of two subsamples from the alder nodule sample gave reasonably replicable readings of 0.40 and 0.45 ppm.
- 2.) According to the standard additions method the accuracy of an analysis is indicated by the degree of linearity acheived when absorbance is plotted against standard addition concentration. Four different standard additions were added to aliquots of each sample. In eleven of thirteen samples, the coefficient of correlation was 20.98, in one sample it was 0.95, and in the other 0.87.

RESULTS AND DISCUSSION

The concentrations of molybdenum in leaf-branch samples ranged from less than 0.04 ppm in Acer macrophyllum, Alnus rubra and Taxus brevifolia to 1.02 ppm in Pseudotsuga menziezii (table 1-1). Alnus rubra, despite its dependance on molybdenum for nitrogen-fixation had the lowest concentration.

The concentrations of northwest trees average about 0.40 ppm dry wt. and are similar to the average plant tissue estimate made by Epstein (1965). On the other hand, concentrations for northwestern trees are an order of magnitude lower than the average of northeastern trees of 3.5 ppm (Young and Guinn 1966). Whether this is a function of site is not presently known.

The Douglas fir sample contained a suprisingly high concentration of molybdenum, almost three times higher than

the nearest species. This relative accumulation of molybdenum by Douglas fir may have considerable ecclogical significance. Alder regeneration is often very aggressive on cut over Douglas fir sites. Increased concentrations of molybdenum on the forest floor resulting from years of Douglas fir litterfall may be a factor in promoting aggressive alder growth. At any rate, the relative content of soils underlying various aged Douglas fir stands should be investigated.

red alder ranged from 0.01 ppm dry wt. for leaves and branches

The distribution of molybdenum was such that roots
to 2.02 for one seed sample. The distribution in requestion of the distribution of molybdenum was such that roots in the distribution in requestion of the distribution in requestion of the distribution in requestion in the distribution in the distribution in requestion in the distribution in the

The molybdenum concentration in nodules is approximately double the highest concentration in any vegetative tissue. This suggests that nodules have a capacity to concentrate the element. This is to be expected because nitrogenase, located in the nodule, requires molybdenum.

The most suprising finding was the high concentration of Mo in red alder seeds, 2.02 ppm dry wt. in northern California seeds and 1.33 in local seeds. This concentration is approximately 15% the average for vegetative tissue and 4% that of nodules. Similarly Lavy and Barber (1963) observed rolybdenum accumulation in the seeds of soy beans. Investigations of this phenomena should be carried out to determine whether a physical phenomena of a capacital adaptation.

The relatively high concentrations in seeds may be important in the ecological strategy of red alder. A stockpile of molybdenum in alder seeds may allow rapid establishment of the seedlings nitrogen fixing capacity independent, initially, of soil concentrations.

Many important questions remain unanswered and deserve discussion and research, especially that of the interactions of pH, Mo availability and growth response. It has been shown that alder actually acidifies the soil down to the soil pH range of 3.8-4.8, approximately one pH unit lower than conifers (Ugolini 1968). Contrary to what one might expect, mature red alder seems to lack nitrogen or molybdenum deficiency symptoms. This suggests that the trees are able to obtain an adequate supply. Perhaps Mo becomes available through mycorrhizal associations or alder is capable of altering soil chemistry so that Mo becomes available. Neal et al. (1968) state that ectotrophic mycorrhizae on red alder increase nutrient absorbing capacity and alter soil chemistry and microbial populations in the rhizosphere. Hatch (1937) showed that rines with mycorrhiza absorbed significantly higher amounts of phosphorus than other nutrients in relation to non-mycorrhizal pines. The chemical activity and availability of phosphorus in acid soils is similar to molybderum. Mitchell (1964) states that uptake of Mo from acid soils can occur if the organic matter content is high. Presumably organic chelation protects Mo from entry into insoluble compounds. Another possible source of molybderum is from decomposition of carbonaceous plant remains

in anaerobic soils which can lead to increased solubilization of trace element complexes of Zn, Co, Mo, Cr, and Cu on waterlogged land (Ng and Bloomfield 1961, 1962).

LITERATURE CITED

- Anderson, A.J. 1946. Molybdenum in relation to pasture improvement in S. Australia. J. Counc. Sci. Ind. Res., 19:1-15.
- Anderson, A.J. and D.V. Moye. 1952. Lime and Mo in clover development on acid soils. Aust. J. of Agric. Res., 3:95.
- Epstien, E. 1965. Mineral metabolism. in: J. Bonner and Varner.

Plant biochemistry. Academic Press, New York, pp. 438-461.

- Fortescue and Martin. 1970. Micronutrients and forest nutrition.

 in: D.E. Reichle. Analysis of temperate forest eccsystems.

 Springer Verlog.
- Hatch, A.B. 1937. The physical basis of mycotrophy in the genus <u>Finus</u>. Black Rock For. Bull., 6:1-168.
- Jones, L.H.P. 1957. The solubility of Moy in simplified systems and aqueous soil suspensions. J. Soil Sci. 8:312-327.
- response of soybeans to Modapplications and the Modeontent of the seeds produced. agron. J., 55:154-155.
- Meyer, F.H. 1966. Mycorrhiza and other plant symbiosis. in: S.M. Henry. Symbiosis. Academic Fress, New York.
- Mitchell, R. 1964. Trace elements in soils. in: F.E. pear. Chemistry of the soil. Reinhold, New York. pp. 320-368.
- Neal, J.L., J.M. Trappe, K.C. Lu and W.B. bollen, 1968. Some ectotrophic mycorrhiza of Alnus rubra. in: J.M. Trappe et al. Biology of alder. Pac. NW For. and Ran. Exp. Sta. Fortland, Oregon. pp.179-184.

- Ng. S.K. and C. Bloomfield. 1961. The solution of minor element oxides by decomposing plant remains. Geochem. Cosmochim. Acta., 24:206-225.
- Ng, S.K. and C. Bloomfield. 1962. The effect of flooding and aeration on the mobility of certain trace elements in soils. Plant Soil, 16:108-135.
- Rubins, E.J. 1956. Modeficiencies in the U.S. Soil Sci. 81:171.
- Ugolini, F.C. 1968. Soil development and alder invasion in a recently deglaciated area of Glacier Bay, Alaska. in:

 J.M. Trappe et al. Biology of alder. Pac. Nw For. and Ran.

 Exp. Sta. Portland, Oregon. pp.115-140.
- Young, H.E. and V.P. Guinn. 1966. Chemical elements in complete mature trees of seven species in Maine. Tappi., 49:190-197.
- Zavitkovski, J. and M. Newton. 1968. Effect of organic matter and combined nitrogen on nodulation and nitrogen-fixation in red alder. in: J.M. Trappe et al. Biology of alder.

 Pac. NW For. and Ran. Exp. Sta. Portland, Oregon. pp.209-224.

Table 1-1. The concentration of molybdenum in ppm dry weight of leaf-branch samples of six northwestern trees and specific tissues of red alder. (* estimate only)

SAMPLE CO	NCENTRATION
Leaf-Branch Samples	
Alnus rubra	0.01
Acer macrophyllum	0.03
Taxus brevifolia	0.03
Tsuga heterophylla	0.20
Thuja plicata	0.37
Pseudotsuga menziezii	1.02
Alnus rubra Samples	
bark	0.14
branches	0.01
large roots	0.19
leaves	0.01
fresh litter	0.19*
nodules (subsample A)	0.40
nodules (subsample B)	0.45
<pre>seeds I (Calif. sample) seeds II (local sample) trunck wood</pre>	2.02
	1.33
	0.01*
Control	0.041.

^{1.} Data below the control concentration represent analytically insignificant amounts.